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De Novo Whole-Genome Sequencing of Two Pathogenic *Pasteurella multocida* Type D:6 Strains Isolated from Pigs

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ABSTRACT Here, we report the complete genome sequences of *Pasteurella multocida* strains P504190 and P504188/1, which were isolated from the diseased lungs of a sow and her piglet, respectively. Despite the unusual clinical presentation, whole-genome sequence typing revealed both strains to be capsular type D and lipopolysaccharide (LPS) group 6, commonly found in pigs.

Pasteurella multocida is a facultative anaerobic Gram-negative bacterial pathogen with the ability to infect a broad host range. *P. multocida* is responsible for significant economic losses in pigs worldwide, causing porcine atrophic rhinitis and respiratory tract disease, with the majority of isolates recovered from clinical cases being characterized as capsular type A, D, or F (1–3).

We report here the complete genome sequences of *P. multocida* strains P504190 and P504188/1, which were isolated from the lungs of a sow and her stillborn piglet, respectively, both of which exhibited unusual gross lesions consistent with a severe acute necrotizing bacterial-type pneumonia and pleurisy. Isolated strains were cultured on blood agar base number 2 with sheep blood (Thermo Fisher Scientific), and *P. multocida* was identified from colonies by matrix-assisted laser desorption ionization–time of flight mass spectrometry. Single colonies were grown aerobically at 37°C for 3 h in Oxoid nutrient broth (Thermo Fisher Scientific). Genomic DNA (gDNA) was extracted by lysis of bacterial pellets at 37°C in Tris-EDTA buffer containing lysozyme and RNase A and then was incubated at 65°C following the addition of proteinase K and SDS. DNA was purified using SPRI beads (Qiagen) and quantified with the Quant-iT double-stranded DNA high-sensitivity kit (Thermo Fisher Scientific). For Illumina sequencing, gDNA libraries were prepared using the Nextera XT library preparation kit and sequenced using the Illumina NovaSeq platform using a 250-bp paired-end protocol. For Oxford Nanopore Technologies sequencing, gDNA libraries were prepared using the rapid barcoding (SQK-RBK004) kit, loaded in a FLO-MIN106 (R.9.4.1) flow cell, and sequenced using a GridION system, with live base calling (Guppy v4.2.2). Illumina reads were adapter trimmed (Trimmomatic v0.30 with a sliding window quality cutoff value of Q15 [4]), and trimmed raw data analysis was performed on the Galaxy platform (<https://usegalaxy.eu/>) (5). Read quality was assessed using FastQC v0.73+galaxy0 (6) and NanoPlot v1.36.2+galaxy1 (7). Illumina sequencing resulted in 458,547 paired-end reads (average read length, 672 bp) and 800,353 paired-end reads (average read length, 554 bp) for P504190 and P504188/1, respectively. Nanopore sequencing produced 19,992 reads (average read length, 7,246.7 bp; read N_{50} , 17,532 bp) for P504190 and 31,718 reads (average read length, 8,573.0 bp; read N_{50} , 19,471 bp) for P504188/1. *De novo* assembly of Nanopore raw reads was performed using Flye v2.9+galaxy0 (8), followed by polishing with Illumina reads using Pilon v1.20.1 (9) after mapping with BWA-MEM v0.7.17.2 (10). Assembly produced single circular contigs (oriented to gene *dnaA*) of 2,327,408 bp (GC content, 40.31%) and 2,327,355 bp (GC content, 40.31%)

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for P504190 and P504188/1, respectively. The average genome coverage for P504190 Illumina and Nanopore read sequences was 94× and 60×, respectively, and that for P504188/1 sequences was 113× and 167×. Assembly metrics were calculated using QUAST v5.0.2+galaxy1 (11). The complete genomes were annotated using the NCBI Prokaryotic Genome Annotation Pipeline v5.0 (12). Default settings were used throughout for all utilized software packages.

The whole-genome sequences were queried in the *Pasteurella multocida* Bacterial Isolate Genome Sequence Database (<https://ivsmist.sund.ku.dk>), which typed both strains as capsular type D and lipopolysaccharide (LPS) group 6 (13).

Data availability. The genome sequences are available in GenBank/EMBL/DDBJ under accession numbers [CP110621](https://ncbi.nlm.nih.gov/nucl/CP110621) (P504190) and [CP110620](https://ncbi.nlm.nih.gov/nucl/CP110620) (P504188/1). Raw sequence reads are available under BioProject accession number [PRJNA896625](https://ncbi.nlm.nih.gov/bioproject/PRJNA896625).

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REFERENCES

- Peng Z, Wang X, Zhou R, Chen H, Wilson BA, Wu B. 2019. *Pasteurella multocida*: genotypes and genomics. *Microbiol Mol Biol Rev* 83:e00014-19. <https://doi.org/10.1128/MMBR.00014-19>.
- Liu S, Lin L, Yang H, Wu W, Guo L, Zhang Y, Wang F, Wang X, Song W, Hua L, Liang W, Tang X, Chen H, Peng Z, Wu B. 2021. *Pasteurella multocida* capsular: lipopolysaccharide types D:L6 and A:L3 remain to be the main epidemic genotypes of pigs in China. *Anim Dis* 1:26. <https://doi.org/10.1186/s44149-021-00031-7>.
- García N, Fernández-Garayzábal JF, Goyache J, Domínguez L, Vela AI. 2011. Associations between biovar and virulence factor genes in *Pasteurella multocida* isolates from pigs in Spain. *Vet Rec* 169:362. <https://doi.org/10.1136/vr.d4869>.
- Bolger AM, Lohse M, Usadel B. 2014. Trimmomatic: a flexible trimmer for Illumina sequence data. *Bioinformatics* 30:2114–2120. <https://doi.org/10.1093/bioinformatics/btu170>.
- Afgan E, Baker D, van den Beek M, Blankenberg D, Bouvier D, Čech M, Chilton J, Clements D, Coraor N, Eberhard C, Grüning B, Guerler A, Hillman-Jackson J, Von Kuster G, Rasche E, Soranzo N, Turaga N, Taylor J, Nekrutenko A, Goecks J. 2016. The Galaxy platform for accessible, reproducible and collaborative biomedical analyses: 2016 update. *Nucleic Acids Res* 44:W3–W10. <https://doi.org/10.1093/nar/gkw343>.
- Andrews S. 2010. FastQC: a quality control tool for high throughput sequence data. <https://www.bioinformatics.babraham.ac.uk/projects/fastqc>.
- De Coster W, D'Hert S, Schultz DT, Cruts M, Van Broeckhoven C. 2018. NanoPack: visualizing and processing long-read sequencing data. *Bioinformatics* 34:2666–2669. <https://doi.org/10.1093/bioinformatics/bty149>.
- Kolmogorov M, Yuan J, Lin Y, Pevzner PA. 2019. Assembly of long, error-prone reads using repeat graphs. *Nat Biotechnol* 37:540–546. <https://doi.org/10.1038/s41587-019-0072-8>.
- Walker BJ, Abeel T, Shea T, Priest M, Abouelliel A, Sakthikumar S, Cuomo CA, Zeng Q, Wortman J, Young SK, Earl AM. 2014. Pilon: an integrated tool for comprehensive microbial variant detection and genome assembly improvement. *PLoS One* 9:e112963. <https://doi.org/10.1371/journal.pone.0112963>.
- Li H. 2013. Aligning sequence reads, clone sequences and assembly contigs with BWA-MEM. *arXiv* 1303.3997v2. <https://doi.org/10.48550/arXiv.1303.3997>.
- Gurevich A, Saveliev V, Vyahh N, Tesler G. 2013. QUAST: quality assessment tool for genome assemblies. *Bioinformatics* 29:1072–1075. <https://doi.org/10.1093/bioinformatics/btt086>.
- Tatusova T, DiCuccio M, Badretdin A, Chetvernin V, Nawrocki EP, Zaslavsky L, Lomsadze A, Pruitt KD, Borodovsky M, Ostell J. 2016. NCBI Prokaryotic Genome Annotation Pipeline. *Nucleic Acids Res* 44:6614–6624. <https://doi.org/10.1093/nar/gkw569>.
- Christensen H, Sajid SM, Bisgaard M, Magistrali CF, Massacci FR, Liman M, Menke T, Bischoff H, Olsen JE. 2022. Prediction of *Pasteurella multocida* serotypes based on whole genomic sequences. *Vet Microbiol* 271:109492. <https://doi.org/10.1016/j.vetmic.2022.109492>.